PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 : C07F 9/10, A61K 31/685, 35/30 — A1

(11) International Publication Number: WO 93/21190

(43) International Publication Date: 28 October 1993 (28.10.93)

(21) International Application Number: PCT/EP93/00915

(22) International Filing Date: 16 April 1993 (16.04.93)

(30) Priority data:
PD92A000073 17 April 1992 (17.04.92) IT

(71) Applicant *(for all designated States except US)*: FIDIA S.P.A. [IT/IT]; Via Ponte della Fabbrica, 3/A, I-35031 Abano Terme (IT).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)* : DI MARTINO, Alessandro [IT/IT]; Galleria Manzoni, 2, I-35036 Montegrotto Terme (IT). CALLEGARO, Lanfranco [IT/IT]; Via Bravi, 35, I-35020 Ponte di Brenta (IT). TOFFANO, Gino [IT/IT]; Via De Amicis, 2, I-35036 Montegrotto Terme (IT).

(74) Agent: VOSSIUS & PARTNER; Siebertstrasse 4, P.O. Box 86 07 67, D-8000 Munich 86 (DE).

(81) Designated States: AU, BB, BG, BR, CA, CZ, FI, HU, JP, KP, KR, LK, MG, MN, MW, NO, NZ, PL, RO, RU, SD, SK, UA, US, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published
With international search report.

(54) Title: METHOD FOR THE PREPARATION AND PURIFICATION OF PHOSPHOLIPID MIXTURES FREE FROM CONTAMINATION BY UNCONVENTIONAL VIRUSES

(57) Abstract

A phospholipid mixture in which the biological and pharmacological properties are maintained but from which unconventional viruses causing e.g. bovine spongiform encephalopathy are selectively eliminated may be prepared by extraction of phospholipids from bovine brain by a mixture of organic solvents of which one is a chlorinated hydrocarbon or by silica gel chromatography with elution with a succession of eluents consisting of chloroform, lower alcohols and water, or by a combination of extraction and silica gel chromatography. Purified phospholipids can be used in pharmaceutical compositions.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	FR	France	MR	Mauritania
AU	Australia	GA	Gabon	MW	Malawi
BB	Barbados	GB	United Kingdom	NL	Netherlands
BE	Belgium	GN	Guinea	NO	Norway
BF	Burkina Faso	GR	Greece	NZ	New Zealand
BG	Bulgaria	HU	Hungary	PL	Poland
BJ	Benin	IE	Ireland	PT	Portugal
BR	Brazil	IT	Italy	RO	Romania
CA	Canada	JP	Japan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo			SE	Sweden
CH	Switzerland	KR	Republic of Korea	SK	Slovak Republic
CI	Côte d'Ivoire	KZ	Kazakhstan	SN	Senegal
CM	Cameroon	LI	Liechtenstein	SU	Soviet Union
CS	Czechoslovakia	LK	Sri Lanka	TD	Chad
CZ	Czech Republic	LU	Luxembourg	TG	Togo
DE	Germany	MC	Monaco	UA	Ukraine
DK	Denmark	MG	Madagascar	US	United States of America
ES	Spain	ML	Mali	VN	Viet Nam
FI	Finland	MN	Mongolia		

"METHOD FOR THE PREPARATION AND PURIFICATION OF PHOSPHOLIPID MIXTURES FREE FROM CONTAMINATION BY UNCONVENTIONAL VIRUSES"

OBJECT OF THE INVENTION

The present invention relates to a process for the preparation of phospholipid mixtures which selectively eliminates contaminants associated with potentially pathogenic unconventional viruses, without altering the biological and pharmacological characteristics of the mixture.

BACKGROUND OF THE INVENTION

Extracts from bovine brain have been used as therapeutic agents for many years. Of such extracts, purified phospholipids represent the

active principle of a large number of pharmacological specialties. Experimental and clinical documentation of the pharmacological properties of phospholipids have been available for many years (Bruni A. et al., Nature 260, 331, 1976; Hirata F. et al., Nature 275, 219, 1978; Delwaide P.J. et al., Acta Neurol. Scand. 73, 136, 1986).

It has long been known that it is possible to extract, at research level, mixtures of phospholipids (which have not yet been fully characterized) (J. Folch, et al., J. Biol. Chem. 226: 497, 1957; N. Radin, Methods Enzymol. 72: 5, 1981; E.G. Bligh, Can. J. Biochem. Physiol. 37: 911, 1959), but none of the aforesaid methods was developed with a view to demonstrating the elimination and destruction of components associated with unconventional viruses. One reason therefor is that at the time the above mentioned procedure was found such diseases caused by unconventional viruses were not known to affect the mammalian species whose brains were used for extraction. Another reason is that no available method for the identification of potentially dangerous components was available. Pathological conditions may sometimes occur where the pathogenic agent or agents have not been identified. One such pathology, called bovine

spongiform encephalopathy (BSE), was first reported in England in 1986 (Wells G. et al. Vet. Record, 419, 1987).

The disease is so called because of the spongy appearance of the brain tissue of affected animals when observed under a microscope; the main lesions are constituted by extensive intraneuronal vacuolation.

All current evidence indicates that BSE belongs to the group of degenerative encephalopathies of the central nervous system, caused by the family of unconventional, transmissible agents whose outcome is invariably fatal (Fraser et al., Vet. Record 123: 472, 1988; Hope et al., Nature 336: 390, 1988). Such diseases include scrapie of sheep and goats, the chronic wasting disease (CWD) which affects captive mule deer, transmissible mink encephalopathy affecting animals on mink ranches, and three human diseases: kuru, Creutzfeldt-Jakob disease (CJD) and the Gerstmann-Straüssler-Scheinker syndrome (GSS). The histopathological lesions found in brains affected by these diseases are similar to those caused by BSE.

Many theories have been put forward as to the nature of these etiological agents, which are different from any known infectious agent, and therefore known as unconventional agents. Due to

the long incubation period between infection and the onset of clinical symptoms, they are also known as "slow viruses".

Since the first few cases in 1986, the disease has reached epidemic proportions in Britain, affecting over 40,000 head of cattle. Affected animals show no signs of the disease for several years (the incubation time is 4-5 years), but once the symptoms have appeared, the course of the disease is rapid and terminates invariably in death.

The results of an epidemiological study carried out by the Central Veterinary Laboratory of the British Ministry of Agriculture (Wilesmith et al, Vet. Record, 123: 638, 1988) identified the source of infection in concentrated animal foodstuffs made from the carcasses of ruminants and sold as meat and bone meal. As the encephalopathies can be transmitted to a wide range of animal species, it is feasible that BSE is the result of infection by the etiologic agent responsible for scrapie, transmitted to cattle in contaminated foodstuffs (Morgan KL, Vet. Record 122: 445, 1988).

On the basis of the results of these studies, the British government issued an order which came into effect on 18th July 1988, outlawing the sale and supply of animal

foodstuffs containing proteins derived from ruminants.

It is commonly thought that various circumstances contributed to the sudden outbreak of BSE in the United Kingdom (Cherfas J., Science Feb. 1990, 523).

Firstly, the number of sheep in the U.K. increased rapidly in the late 70's and early 80's, and with it the occurrence of scrapie, an endemic disease of the ovine species in Europe for over 250 years (Pattison et al, Vet. Record 90: 465, 1972). At the same time, in the wake of the oil crisis, animal feed production plants switched their methods of production to a low temperature system which was probably less effective in destroying the highly resistant scrapie agent. All meat and bone meal producers except one discontinued the use of organic solvents, such as benzene, hexane and trichloroethylene, to remove excess fat from soybean and bone meal. Perhaps most significant of all was that the final stage of heating the products to remove the solvents was consequently left out: indeed this step required very high temperatures.

Moreover, government policy encouraged breeders to produce more milk, and wean calves early by feeding them protein-enriched diets.

These were often of poor quality, since protein meal made from meat and bone was cheaper than products made with soybean and fish which are more reliable sources of protein.

Studies on how BSE is transmitted are fundamental to research. The most important aspect of these experiments is that, by identifying the limits of the inter-species barriers to transmission of the pathogenic agent, it may be possible to assess the risk of infection by BSE to any one species. Fraser et al (Vet. Record, 123:472, 1988) demonstrated that the disease could be transmitted to mice by inoculating extracts from the brains of cattle which had died from BSE into the brains of mice which subsequently developed the disease. Later, Barlow et al (Vet. Record, 3 Feb. 1990) transmitted the disease to mice by feeding them infected brains. It was the first evidence that BSE could be transmitted by the oral route. No other tissue from infected animals (spleen, spinal cord, lymphatic tissues, milk etc.) has been able to produce the disease in mice. While evidence exists that scrapie can be transmitted from ewe to lamb, there is no proof so far of possible vertical or horizontal transmission of the etiological agent of BSE among cattle.

The agents which cause subacute spongiform encephalopathies are extremely resistant to standard decontamination procedures. Currently available data on this aspect mainly originate from studies on the inactivation of scrapie and Creutzfeldt-Jakob disease agents.

The etiological agent of scrapie is highly resistant to temperature sterilization. Prolonged exposure to temperatures of up to 80°C only slightly reduces their infectivity; higher temperatures however markedly reduce infectivity (Hunter et al, J. Gen. Microbiol. 37: 251,1964). A small quantity of infectious "virus" remains viable when suspensions of infected material are heated to 100°C for 1 hour or to 118°C for 10 minutes. Recently the need was felt to update standards of sterilizing these infectious agents by steam autoclaving. Current methods of autoclaving in the United States for decontamination from Creutzfeldt-Jakob disease involve treatment at 132°C for 1 hour (Rosenberg et al, Annals of Neurology 19: 75, 1986), and is based on studies carried out on brain homogenates containing scrapie or Creutzfeldt-Jakob agents (Brown et al, J. of infectious diseases 153: 1145, 1986). In the U.K., current standards of autoclaving for decontamination from Creutzfeldt-Jakob disease involve treatment in an autoclave

at 134-138°C for 18 minutes, and are based on various studies including one by Kimberlin (Kimberlin et al, Journal of Neurological Sciences 59: 355, 1983). Unfortunately, the spongiform encephalopathy agents are very resistant even to common chemical and physical treatments. Solvents such as benzene, hexane, petrol and trichloroethylene have been used as extraction solvents, but little is known of their effects on infectivity. Limited data are available on the chemical inactivation of the infectious agents, mainly because studies require large numbers of animals and long monitoring times.

Concentrations of 0.3% - 2.5% of sodium hypochlorite greatly reduced infectivity in the biological assays, but did not always eliminate it completely (Walker et al., Am. J. Publ. Health 73:661, 1983). Data regarding treatment with up to 0.25 N sodium hydroxide are inconsistent; however at a concentration of over 1 N it appears to be the most efficacious chemical agent of all those studied. Treatment with 6M-8M urea was also reported to be highly variable.

The results of the studies on decontamination thus show that, although most of the infectivity is quickly destroyed by many of the different physical and chemical treatments, the

existence of small subpopulations of infectious agents resistant to inactivation makes complete sterilization of contaminated materials extremely difficult in practice.

Once BSE had been identified as a "scrapie-like" disease, important epidemiological and analytical questions were raised, the latter in particular being aimed at identifying the agent that could be associated with the infection.

The sudden appearance of BSE and all the other aspects of these neurological disorders still to be clarified have caused necessary consideration to be given to the problem, especially by those involved in the preparation of products deriving from bovine material.

Before BSE ever occurred it was obvious that the state of the art required a product to be pharmaceutically acceptable and free from those biological contaminants which were known at the time to constitute a potential risk to health. But clearly, the subsequent appearance of the above-described pathology in adult cattle has made it necessary to obtain an active principle which, without losing its known therapeutic properties, is characterized by the complete absence of unconventional viral agents, to be achieved by the use of specific procedures to guarantee the inactivation of these unconven-

tional viral agents and the complete elimination of infectivity, and to use specific methodologies by which to identify such agents.

Indeed, it may not be enough to use raw material which has been certified as suitable for consumption, to obtain compounds or mixtures of the same for pharmaceutical purposes. Obviously, the elimination of infectivity, in this case BSE, must be assessed by analyzing its biological action *in vivo*. This would serve as a test of the various steps of the process, but not as an overall check of the same. The elimination of infectivity associable with bovine spongiform encephalopathy is preferably assessed by means of experiments involving spiking with infective agents. It is especially preferred that the analysis of destruction of the infective agent introduced by spiking is performed *in vivo*. This analysis of the biological action *in vivo* is necessary since scientists do not agree over associating the infection with certain proteins or their fragments attributable to the unconventional infective agents such as the proteins PrP^{sc} or PrP_{27-30} associated with BSE. Clearly, the extraction process which eliminates infectivity must at the same time leave the biological activity of the active principle intact, since this is essential for its

therapeutic use. (Committee for Proprietary Medicinal Product. Ad hoc working party on biotechnology-pharmacy; Biologicals 19: 247, 1991). Scientific research has produced, on the one hand, methods which guarantee suitable mixtures of phospholipids or their single fractions to be obtained devoid of protein, chemical and biological contaminants. On the other hand, methods with demonstrated efficacy in destroying infectivity associated with slow viruses are known, but no method exists by which it is possible to obtain, on an industrial scale, a product, as desired, in pure, pharmacologically active form, free from infectivity associable with pathogenic agents definable as slow viruses.

BRIEF DESCRIPTION OF THE INVENTION

The purpose of the present invention is to provide a product in pharmacologically active form and, at the same time, being characterized by the absence of slow viruses. Such a product can be obtained by an advantageous process which can be applied to industrial production. The process offers innovative qualities founded on the suitable sequencing of its various extraction steps. During the various steps of this process, infective contaminants associable with slow viruses such as the bovine spongiform

encephalopathy agent are eliminated, while the activity of the mixture, which represents the therapeutic activity of the product itself, remains unaltered. The product deriving from this process is constituted by a definite mixture of phospholipids or single fractions obtained from bovine brain or parts of the same.

DETAILED DESCRIPTION OF THE INVENTION

According to the present invention, a phospholipid mixture which is essentially free from infective components while maintaining the pharmacological properties of said phospholipid mixture is prepared by a process comprising subjecting a material derived from nervous tissue a purification step selected from the group consisting of

A) extracting said material with an organic solvent,

B) loading a suspension of said material on a silica gel column and eluting said column with a mixture of organic and aqueous solvents,

and a combination of steps A) and B).

The nervous tissue is preferably derived from bovine brain.

In a preferred variant of step A) said nervous tissue material is subjected to an extraction with a solvent mixture including a

chlorinated hydrocarbon to obtain a first raw phospholipid-comprising mixture, subjecting said first raw phospholipid-comprising mixture a further purification step, and isolating the further purified product.

According to the invention, it is preferred that the suspension of the phospholipid mixture to be loaded on a silica gel column has been prepared by a method comprising the steps of

a) extracting bovine brain to obtain a crude extract comprising phospholipids,

b) filtering said crude extract to obtain a filtrate,

c) adding a precipitating agent to said filtrate to obtain a precipitate,

d) isolating said precipitate to obtain said first raw phospholipid-comprising mixture,

e) purifying said first raw mixture to obtain a second phospholipid-comprising mixture,

f) suspending said second mixture in a liquid.

The extraction in step a) is conducted with a mixture of organic solvents such as acetone in combination with chlorinated hydrocarbons, especially 1,1,1-trichloroethane. The extraction

is performed under vigorous stirring for an extended period of time, e.g. 30-60 minutes. The temperature is preferably room temperature. After the extraction process which results in a suspension, the suspension is centrifuged, whereby insoluble material and the upper acetone-comprising phase were discarded. The crude extract is then filtered according to step b). In step c) a precipitating agent such as acetone is added to the filtrate; the mixture is stirred for about 30-60 minutes at a temperature below room temperature, e.g. at 13-17°C, preferably at 15°C. Thereafter the precipitate is isolated in step d) by means of centrifugation. The isolated product is the first raw material (phospholipid-comprising mixture). Said first raw material is redissolved in a solvent mixture comprising a chlorinated hydrocarbon such as 1,1,1-trichloroethane in admixture with an alcohol such as absolute ethanol. Thereby the first raw material is solubilized. The purification in step e) of the first raw material is performed by the addition of water, shaking the mixture for a period of 30-90 minutes, preferably 60 minutes, at room temperature. Separation of the phases is then performed by centrifugation. The process of solubilization and precipitation and centrifugation may be repeated. When the organic phase

containing the phospholipid material has been isolated it is precipitated by means of e.g. acetone, and the mixture is left standing for 30-60 minutes at a temperature of 13-17°C, preferably 15°C. The precipitate is isolated by centrifugation and may be further rinsed with acetone. The precipitate is dried, e.g. in vacuum, to yield a second raw material (a second phospholipid-comprising mixture). This second raw material may be further purified by the addition of a saline to a suspension of the second raw material in a chlorinated hydrocarbon such as chloroform in admixture with an alcohol such as methanol. The mixture of aqueous and non-aqueous solvents comprising the second raw material is then shaken for 15-30 minutes at room temperature. Centrifugation of the two phases and recovery of the organic phase yields a solution of the desired phospholipid mixture. The phospholipids can be precipitated by means of acetone; in order to ensure full precipitation the mixture may be left standing for a period of time of 30-60 minutes at a temperature below room temperature, e.g. at 13-17°C, preferably 15°C. Centrifugation, rinsing with acetone and drying yield the final product. The final product may be sterilized by being exposed to elevated temperature.

The alternative purification method for purification consists of loading the material derived from nervous tissue on a silica gel column. The column is pre-washed with a chlorinated hydrocarbon such as chloroform before the nervous tissue derived material is loaded thereon. The elution of the desired phospholipid mixture is performed with the chlorinated hydrocarbon, preferably chloroform, to which is added one or more alcohols and water. The alcohol is selected from the group consisting of methanol, ethanol, propanols, butanols and pentanols. The elution is performed with eluents in e.g. the following sequence: chloroform, chloroform/ethanol, chloroform/methanol/water (4%) and chloroform/methanol/water (12.5%). The flow is divided in fractions which are analyzed for the content of phospholipids. Relevant fractions are pooled, and the phospholipids are recovered and dried.

When a combination of extraction and column chromatography is used in the purification process according to the invention, the second raw material from step e) is suspended in a suitable liquid comprising a chlorinated hydrocarbon such as chloroform and an alcohol such as methanol. The suspension is loaded on a silica gel column and eluted as described above.

After the elution the desired end product is recovered from the eluting solvents, preferably by means of vacuum drying.

An especially preferred process variant consists of extraction of phospholipids from bovine brain by a mixture of chlorinated solvents, for example 1,1,1-trichloroethane, and acetone, at room temperature and for at least 30 minutes; or silica gel chromatography of a phospholipid mixture with elution with a succession of suitable mixtures of chloroform, ethanol, methanol and water; or a combination of the above procedures.

In a preferred variant of the process according to the invention the product is a phospholipid mixture comprising at least two components selected from the group consisting of phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, plasmalogen choline, plasmalogen ethanolamine, plasmalogen serine, phosphatidic acid, diphosphoinositide and sphingomyelin. In still another variant, the phospholipid preparable by the invention comprises only a single phospholipid fraction such as phosphatidylserine.

MATERIALS AND METHODS

The bovine brains used in the process of extraction of the phospholipid mixture showed, on histological analysis, the fibrils typical for tissues belonging to materials from animals with the infection.

BIOLOGICAL TEST FOR INFECTION

The animals used in these experiments were Golden Syrian hamsters (LVG/Lak). Tests for infection were carried out on groups of four female weanlings which had received intracerebral (i.c.) inoculation with 0.05 ml of the samples diluted ten times in sterile PBS. The intracerebral inoculations were effected by trained staff using disposable glass syringes with 26G, 3/8-inch sterile needles.

The final, sterilized product, concentrated 10 times, was used entirely.

The animals were examined twice a week or more for a period of 12 months, for signs of the characteristic, clinical, neurological symptoms. The onset of early symptoms in each animal was recorded, and the animals were sacrificed when the disease was in its final stage. Their brains were cut into halves, one fixed in 10% formalin and the other stored at -70°C. Histopathological examination was made in all animals which died of

suspect causes and those which had shown signs of neurological disorders. At the end of the observation time, all surviving animals were sacrificed and a histopathological assessment was made on their brains.

The infective titer was calculated at the "final end point" according to the method of Reed and Munch, and is expressed as log LD_{50}/ml.

Brief description of Figure 1

Figure 1 illustrates an example of the compositions of the phospholipid mixture purified as described.

FIGURE 1

Example of silica gel chromatography of the following samples:

Lane 1: standard phospholipid phosphatidylethanolamine (PE)

Lane 2: standard phospholipid phosphatidic acid (PA)

Lane 3: standard phospholipid phosphatidylinositol (PI)

Lane 4: standard phospholipid phosphatidylserine (PS)

Lane 5: standard phospholipid phosphatidylcholine (PC)

Lane 6: standard phospholipid sphingomyelin (SM)

Lane 7: final product (phospholipid mixture)

The phospholipid mixture, obtained as described and free from contaminants associated with unconventional, potentially dangerous infective agents, can also be used for the preparation of single components of the phospholipid mixture, such as phosphatidylserine.

Thanks to its known pharmacological properties (G. Toffano et al., Pharmacol. Res. Comm. 8: 581, 1976; Bruni A. et al. Adv. Exp. Med. Biol. 72, 271-287, 1976; A. Leon et al., J. Neurochem. 30: 23, 1978; E. Boarato et al., Agents and actions 14: 613, 1984; Ponzin D. et al.: Immunopharm. 18, 167, 1989; Monastra G. et al. Lymphokine and cytokine Res. 11, 1, 39, 1992), the phospholipid mixture (or its single components), prepared by the process of the invention, can be used in general to prepare pharmaceutical compositions which are efficacious in numerous pathologies (with different etiopathogenic causes) in particular of the central nervous system and the immune system, especially degenerative pathologies associated with immune dysfunctions. The following can be cited: psychoorganic syndromes characterized by

involution or cerebrovascular insufficiencies, behavioral impairments correlated with neuroendocrine alterations, depressive syndromes, anxious-depressive states, degenerative pathologies also associated with immune dysfunctions, multiple sclerosis, Alzheimer's disease, amyotrophic lateral sclerosis, encephalitis of various origin, and in pathologies associated with alterations in intestinal absorption of the fatty acids.

The pharmaceutical compositions according to the present invention can be administered by oral or parenteral route, preferably by intramuscular route, by intravenous injection or by infusion. The preparations can be as solutions of the compound (or freeze-dried powders of the compound) in association with one or more pharmaceutically acceptable vectors or diluents and contained in buffered media with a suitable pH and isotonic with the physiological fluids. Safe storage of the pharmaceutical can be ensured by packing it in vials, ampoules, capsules, single dose packets, as described in the following examples of pharmaceutical preparations. For its therapeutic and possibly also preventive application by said parenteral or oral routes, the dose of active substance depends on the desired effects and on the chosen route of

administration and can be between 0.05 and 12 mg/kg body weight of the patient per dose, corresponding to a daily administration of between about 5 and 2000 mg to an adult human.

Another aspect of the present invention relates to a method for the treatment of pathologies of the central nervous system and degenerative pathologies also associated with immune system dysfunctions comprising the administration of an effective amount of a phospholipid mixture prepared by the present invented process, to a patient in need therefor.

A mixture of phospholipids prepared according to the invention may have the following composition: 30-50% phosphatidylcholine, 24-44% phosphatidylethanolamine, 7-13% phosphatidyl-serine, and 11-27% phosphatidylinositol and minor phospholipids.

When the phospholipids are presented in pharmaceutical preparations, such preparations may have the following composition (expressed in mcg of phosphorus, per 50 mg of mixture): 280-420 mcg P as phosphatidylcholine, 140-210 mcg P as phosphatidylethanolamine, 140-210 mcg P as phosphatidylserine, 8-12 mcg P as plasmalogen choline, 68-102 mcg P as plasmalogen ethanolamine, 40-60 mcg P as plasmalogen serine,

10-24 mcg P as phosphatidic acid, 28-42 mcg P as diphosphoinositide and 68-102 mcg P as sphingomyelin.

Another composition of a mixture of phospholipids prepared according to the invention can be as follows (expressed in mcg of phosphorous per 100 mcg of total phosphorous): 25-45 mcg phosphatidylcholine, 12-22 mcg P as phosphatidylethanolamine, 12-22 mcg P as phosphatidylserine, 0.5-1.5 mcg P as plasmalogen choline, 5-13 mcg P as plasmalogen ethanolamine, 2-8 mcg P as plasmalogen serine, 1-3 mcg P as phosphatidic acid, 2-6 mcg P as diphospho-inositide and 5-13 mcg P as sphingomyelin.

Hereafter, for illustrative and not limitative purposes, examples 1-3 describe preparations made from infected bovine brains where the spongiform encephalopathy form was encountered or from protein raw materials obtained from uninfected bovine brains to which constant amounts of infected material from the 263K scrapie strain are added. Examples 4-18 illustrate pharmaceutical compositions with the purified phospholipids. The invention being thus described, it is clear that these methods can be modified in various ways. These modifications are not to be considered as deviations from the spirit and the purpose of the invention and all

those modifications which would appear evident to one skilled in the art are comprised within the scope of the subsequent claims.

EXAMPLE 1

1000 grams of infected frozen bovine brain, were ground to a fine powder. A small aliquot of powder was removed for the infectivity test and stored at -70°C.

All of the powder was dispersed in 900 ml of acetone for 15-30 minutes at room temperature. To this was added 1,1,1-trichloroethane to obtain a weight/volume ratio of 1:3. The suspension was left under vigorous magnetic stirring for 30 minutes at room temperature. It was centrifuged for 20 minutes at 1,000 rpm at 20°C and 500 ml of clear, lower, organic phase was recovered, discarding the upper, acetone/water phase and all insoluble material. It was subsequently left for 60 minutes in a thermostatic bath at a temperature of 7.5°C; it was then filtered through a Gooch funnel (pore size No. 3) to eliminate unsolubilized material. 1750 ml of acetone was added and the mixture was magnetically stirred for 30 minutes at a temperature of 15°C, and it was centrifuged for 10 minutes at 2000 rpm at 15°C. The precipitate (Raw Material 1) was repeatedly washed with 1000

ml of acetone and vacuum-dried at 15°C. 30 g of raw Material 1 was redissolved in 240 ml of a mixture of 1,1,1-trichloroethane and absolute ethanol in the ratio 5:2. Once solubilization was complete, 50 ml of distilled water was added and it was partitioned by shaking for 60 minutes at room temperature. It was then centrifuged at 1000 rpm until complete separation and clarification of the two phases, and the lower, organic phase was recovered. This operation was then repeated for a second time in the same conditions. A small aliquot was removed for biological assay and 720 ml of acetone was added to both these fractions. It was left for 30 minutes at a temperature of 15°C, then centrifuged again at 2000 rpm for 10 minutes. The precipitate was repeatedly washed with 500 ml of acetone. The product was vacuum-dried at 25°C (Raw Material 2). 2.4 g of raw Material 2 was resuspended in 300 ml of chloroform/methanol. To this was added 90 ml of an aqueous 0.4 molar solution of sodium chloride and it was partitioned by shaking for 30 minutes at room temperature. Finally the material was centrifuged at 1000 rpm till complete separation and clarification of the two phases, and the lower, organic phase was recovered. This operation was then repeated a second time in the

same way. The final volume (180 ml) was divided into two identical aliquots to each of which was added the same amount of acetone, namely 540 ml. This was left to react for 30 minutes at a temperature of 15°C, and then centrifuged again at 2000 rpm. The precipitate was washed repeatedly with 500 ml of acetone. The product (10+10 g) was then vacuum-dried at 25°C and recovered in PBS (final product). An aliquot was sterilized at 121°C for 15 minutes then rapidly cooled at room temperature (final, sterilized product). Yield 2%.

The samples for biological assay were recovered in sterile PBS in the following volumes:

Powdered, infected brain	ml 0.15 dilution 10^0
Intermediate product (Raw Material)2	ml 0.33 dilution 10^0
Final product	ml 1.41 dilution 10^1
Final, sterilized product	ml 1.41 dilution 10^1

(10^0 means undiluted; 10^1 means diluted 10 times.)

Table 1 reports an example of the results obtained by assay of the biological activity.

TABLE 1

Sample Dilution	33% Brain Homogenate	Sick Animals	Raw M.2	Sick Animals	Finished Product	Sick Animals	Finished Sterilized Product	Sick Animals
10x conc.					16-21172	0/10	16-21176	0/12
Undiluted			4-21169	0/2	32-21173/74 (ic) 2-21175 (ip)	0/20 0/1	80-2117-80(ic) 4-21181 (ip)	0/50 0/2
10^{-1}			4-21169	0/3				
10^{-2}			4-21170	0/2				
10^{-3}			4-21171	0/1				
10^{-4}	4-21162	3/3						
10^{-5}	4-21163	4/4						
10^{-6}	4-2164	4/4						
10^{-7}	4-21165	4/4						
10^{-8}	4-21166	0/1						
10^{-9}	4-21167	0/4						
TOTAL	24	15/20	16	0/8	50	0/31	100	0/63

Included in the study were all animals that did show clinical signs of scrapie. Animals that died of causes other than scrapie were not included. The column marked "sample" reports the number of animals inoculated at the start of the experiment and the cage number relative to each different sample and dilution. ??? The column marked "sick animals" reports the number of animals with clinical signs of scrapie/number of animals inoculated minus the number of animals that died of causes other than scrapie.

EXAMPLE 2

1000 g of infected frozen bovine brain were ground to a fine powder. A small aliquot was then removed and stored at -70°C for the infectivity assay.

All of the powder was dispersed in 900 ml of acetone for 15-30 minutes at room temperature. To it was added ml of 1,1,1-trichloroethane to obtain a weight/volume ratio of 1:3. The suspension was left under vigorous magnetic stirring for 30 minutes at room temperature. It was centrifuged for 20 minutes at 1000 x rpm at 20°C and the clear, lower, organic phase was recovered (500 ml), discarding the upper, acetone/water phase and all insoluble material. It was then left for 60 minutes, in a thermostatic bath at a temperature of 7.5°C; it was then filtered through a Gooch funnel, pore size 3, to eliminate the insoluble material. 1750 ml of acetone was added and it was left under magnetic stirring for about 30 minutes at a temperature of 15°C, and it was centrifuged for 10 minutes at 2000 x rpm at +15°C. The precipitate (First Raw Material 1) was repeatedly washed with 1000 ml of acetone and left to vacuum-dry at 15°C. 30 g of raw Material 1 was redissolved in 240 ml of a mixture of 1,1,1-trichloroethane/absolute ethanol in the

ratio 5:2. Once solubilization was complete, 50 ml of distilled water was added and it was partitioned by shaking for 60 minutes at room temperature. It was then centrifuged at 1000 rpm until complete separation and clarification of the two phases, and the lower, organic phase was recovered. This operation was repeated a second time in the same way. A small aliquot was removed for biological assay and 720 ml acetone was added to both of these fractions. It was left for 30 minutes at a temperature of 15°C, then centrifuged again at 2000 rpm for 10 minutes. The precipitate was washed repeatedly with 500 ml of acetone. The product was vacuum-dried at 25°C (Second Raw Material 2). The precipitate was repeatedly washed with acetone and resuspended in 300 ml of chloroform/methanol. The material was then loaded onto a silica gel column (6-35 μ) equilibrated in chloroform. The column was eluted with chloroform, chloroform/ethanol in the ratio 7:3, chloroform/methanol/water (65:25:4) and chloroform/methanol/water (50:50:12.5) (12.5 parts of water per volume) in turn. Each fraction was recovered for analysis by TLC chromatography (V.P. Skipski et al., Methods Enzymol., 14, 1969: 530; F, Vitiello et al., J. Chromatogr., 166, 1978: 637). After analysis the

fractions eluted with chloroform/methanol/water (65:25:4) (4 parts of water per volume), corresponding to fractions 12-18, were recovered and vacuum-dried. They were then resuspended by sonication in PBS.

The samples for biological assay were recovered in sterile PBS in the following volumes:

Final product	ml 0.8	dilution 10^1
Final, sterilized product	ml 0.8	dilution 10^1

Table 2 reports an example of the results obtained by assay of the biological activity.

TABLE 2

Sample Dilution	33% Brain Homogenate	Sick Animals	Finished Product	Sick Animals	Finished Sterilized Product	Sick Animals
10x conc.			8-21231	0/5	8-21234	0/5
Undiluted			40-21232/33 (1c)	0/26	40-21235/36(1c)	0/28
10^{-1}						
10^{-2}						
10^{-3}						
10^{-4}	4-21162	3/3				
10^{-5}	4-21163	4/4				
10^{-6}	4-21164	4/4				
10^{-7}	4-21165	4/4				
10^{-8}	4-21166	0/1				
10^{-9}	4-21167	0/4				
TOTAL	24	15/20	48	0/31	48	0/33

Included in the study were all animals that showed clinical signs of scrapie. Animals that died of causes other than scrapie were not included in the study. The column marked "sample" reports the number of animals inoculated at the start of the experiment and the cage number relative to each different sample and dilution. The column marked "sick animals" reports the number of animals with clinical signs of scrapie/number of animals inoculated minus the number of animals that died of causes other than scrapie.

EXAMPLE 3

5 g of silica gel (6-35 μ) were resuspended in a volume of 10-20 ml of chloroform for 5-15 minutes. The gel was loaded onto a vacuum-packed chromatographic column. The column was then washed thoroughly with chloroform. 50 mg of uninfected bovine phospholipids (prepared as described in Example 1; corresponds to "Raw Material 2") were resuspended in 0.5-1.0 ml of chloroform/methanol and a 25% (w/v) homogenate of infected hamster brain 263K was added. This homogenate had a infectivity titer of 9.2 log LD50/ml (Di Martino et al, Arch. Virol., 124, 1992: 111). An identical amount of infected material was added in the same volume of PBS (infected homogenate).

The sample was well shaken to disperse the aqueous phase and 10 μl were taken for biological assay (starting material). The sample was then loaded into a column where it was eluted with the following solvents and their mixtures: chloroform, chloroform/ethanol, (7:3) chloroform/methanol/water, (65:25:4) and chloroform/methanol/water (50:50:12.5) (12.5 parts of water in the organic phase volume).

The rate of flow through the column was between 3 and 10 ml/min, preferably 6 ml min, and fractions of 2 ml each were taken.

The fractions were recovered as follows:

Fraction 1-8 Column front, cholesterol esters, cerebrosides

Fraction 9-11 Sulfatides and traces of phosphatidylcholine

Fraction 12-18 phospholipids

Fraction 19-22 Hydrophilic products and traces of sphingomyelin.

Each fraction was withdrawn for TLC chromatographic analysis (J.C. Touchstone et al., J. Liq. Chromatogr. 6: 179, 1983; F, Vitiello et al., J. Chromatogr., 166, 1978: 637). After analysis the fractions eluted with chloroform/ methanol/water (4 parts of water per volume), corresponding to fractions 12-18, were recovered and vacuum-dried. They were then resuspended by sonication in PBS.

Table 3 reports an example of the results obtained by assay of biological activity.

Sample	Inoculated Animals	Infectivity	% of Infected animals	Mean incubation time(days)
Homogenate	12	yes	100	67 ± 9
Starting Material	12	yes	66	153 ± 39
Fractions 1-8	12	no	0	0
Fractions 9-11	12	no	0	0
Fractions 12-18	12	no	0	0
Fractions 19-22	12	yes	16.7*	182 ± 0*

*Values calculated on one animal only

Included in the study were all animals that showed clinical signs of scrapie. Animals that died of causes other than scrapie were not included.

EXAMPLE 4

One 2 ml ampoule is composed as follows:

hypothalamic phospholipid liposomes corresponding to micrograms of lipidic phosphorous 400

- phosphatidylcholine 40%
- phosphatidylethanolamine + phosphatidylserine 34%
- sphingomyelin 10%
- other phospholipids present in small quantities (phosphatidylinositol, diphosphoinositide, phosphatidic acid, lysolecithin, lysocephalin) 16%

Other components:

- lidocaine hydrochloride 2 mg
- esters of p-oxybenzoic acid 1.2 mg
- monobasic sodium phosphate $2H_2O$ 2.14 mg
- dibasic sodium phosphate 12 H_2O 2.26 mg
- water for injection q.s. ad 2 ml

EXAMPLE 5

One 2-ml ampoule contains:

hypothalamic phospholipid liposomes corresponding to micrograms of lipidic phosphorous 1000

- phosphatidylcholine 40%
- phosphatidylethanolamine + phosphatidylserine 34%
- sphingomyelin 10%

- other phospholipids present in small quantities (phosphatidylinositol, diphosphoinositide, phosphatidic acid, lysolecithin, lysocephalin) 16%

Other components:

- esters of p-oxybenzoic acid 1.2 mg
- monobasic sodium phosphate $2H_2O$ 2.14 mg
- dibasic sodium phosphate 12 H_2O 2.26 mg
- mannitol 100 mg
- water for injection q.s. ad 2 ml

EXAMPLE 6

One 2-ml ampoule contains:

- phosphatidylcholine 140 mcg of P
- phosphatidylethanolamine 70 mcg of P
- phosphatidylserine 70 mcg of P
- plasmalogen-choline 4 mcg of P
- plasmalogen-ethanolamine 35 mcg of P
- plasmalogen-serine 20 mcg of P
- phosphatidic acid 8 mcg of P
- diphosphoinositide 15 mcg of P
- sphingomyelin 35 mcg of P
- Cyanocobalamin 1000 mcg

Other components:

- alpha-diethylamino-2-6 dimethylacetanilideHCl 2.4 mg
- esters of p-oxybenzoic acid 1.2 mg

- monobasic sodium phosphate $2H_2O$ 2.14 mg
- dibasic sodium phosphate $2H_2O$ 2.26 mg
- water for injection q.s. ad 2 ml

EXAMPLE 7

One capsule contains:

- phospholipids from cortical grey matter 50 mg

constituted by:

- phosphatidylcholine 350 mcg of P
- phosphatidylethanolamine 175 mcg of P
- phosphatidylserine 175 mcg of P
- plasmalogen-choline 10 mcg of P
- plasmalogen-ethanolamine 85 mcg of P
- plasmalogen-serine 50 mcg of P
- phosphatidic acid 20 mcg of P
- diphosphoinositide 35 mcg of P
- sphingomyelin 85 mcg of P
- pyridoxine hydrochloride 150 mg
- cyanocobalamin 250 mcg

Other components:

- vegetable oil F.U. 200 mg

The outer shell is constituted by:

- gelatin F.U. 140 mg
- glycerin F.U. 45 mg
- sodium ethyl paraoxybenzoate 0.6 mg
- sodium propyl paraoxybenzoate 0.3 mg
- ethyl vanillin 0.5 mg

- titanium dioxide 2.1 mg
- erythrosine (E127) 2.6 mg

EXAMPLE 8

One 2-ml ampoule contains:

- phosphatidylserine 50.00 mg

Other components:

- lecithin 15.00 mg
- mannitol 100.00 mg
- dibasic sodium phosphate-12H_2O 2.26 mg
- monobasic sodium phosphate-2H_2O 2.14 mg
- water for injection q.s. ad 2 ml

EXAMPLE 9

One 2-ml ampoule contains:

- phosphatidylserine 50.00 mg

Other components:

- mannitol 100.00 mg
- dibasic sodium phosphate-12H_2O 2.26 mg
- monobasic sodium phosphate-2H_2O 2.14 mg
- water for injection q.s. ad 2 ml

EXAMPLE 10

One 10-ml vial in which each ml contains:

- phosphatidylserine 25.00 mg

Other components:

- lecithin 7.50 mg
- mannitol 50.00 mg
- dibasic sodium phosphate-12H_2O 1.13 mg
- monobasic sodium phosphate-2H_2O 1.07 mg
- water for injection q.s. ad 1 ml

EXAMPLE 11

One 10-ml vial in which each ml contains:

- phosphatidylserine 25.00 mg

Other components:

- mannitol 50.00 mg
- dibasic sodium phosphate-12H_2O 1.13 mg
- monobasic sodium phosphate-2H_2O 1.07 mg
- water for injection q.s. ad 1 ml

EXAMPLE 12

One capsule contains:

- phosphatidylserine 100.00 mg

Other components:

- lecithin 30.00 mg
- vegetable oil 270.00 mg

The outer shell is constituted by:

- gelatin 129.00 mg
- glycerol 49.00 mg

-	rust brown E 172	1.10 mg
-	rust red E 172	0.40 mg
-	sodium ethyl p-hydroxybenzoate	0.20 mg
-	sodium propyl p-hydroxybenzoate	0.10 mg

EXAMPLE 13

One capsule contains:

-	phosphatidylserine	100.00 mg

Other components:

-	vegetable oil	270.00 mg

The outer shell is constituted by:

-	gelatin	129.00 mg
-	glycerol	49.00 mg
-	rust brown E 172	1.10 mg
-	rust red E 172	0.40 mg
-	sodium ethyl p-hydroxybenzoate	0.20 mg
-	sodium propyl p-hydroxybenzoate	0.10 mg

EXAMPLE 14

One vial complete with ampoule of solvent for parenteral administration, each vial containing:

Freeze-dried active component

-	phosphatidylserine	50.00 mg

Other components:

-	lecithin	15.00 mg
-	mannitol	100.00 mg

- water for injection q.s. ad 4 ml

EXAMPLE 15

One vial complete with ampoule of solvent for parenteral administration, each vial containing:

Freeze-dried active component

- phosphatidylserine 50.00 mg

Other components:

- mannitol 100.00 mg
- water for injection q.s. ad 4 ml

EXAMPLE 16

One single-dose pack (granules for oral use to mix with water before use) contains:

- phosphatidylserine 100.00 mg

Other components:

- sodium ascorbate 5.00 mg
- aspartame 5.00 mg
- colloidal silica 10.00 mg
- soybean lecithin 30.00 mg
- natural flavoring 40.00 mg
- mannitol 350.00 mg
- fructose q.s. ad 1.50 g

EXAMPLE 17

One single-dose pack (granules for oral use to mix with water before use) contains:

- phosphatidylserine 100.00 mg

Other components:

- sodium ascorbate	5.00 mg
- aspartame	5.00 mg
- colloidal silica	10.00 mg
- natural flavoring	40.00 mg
- mannitol	350.00 mg
- fructose	q.s. ad 1.50 g

EXAMPLE 18

One single-dose pack (granules for oral use to mix with water before use) contains:

- phosphatidylserine	200.00 mg

Other components:

- sodium ascorbate	10.00 mg
- aspartame	20.00 mg
- colloidal silica	20.00 mg
- soybean lecithin	60.00 mg
- natural flavoring	80.00 mg
- mannitol	700.00 mg
- fructose	q.s. ad 3.00 g

CLAIMS

1. A process for the preparation of a phospholipid mixture which is essentially free from infective components while maintaining the pharmacological properties of said phospholipid mixture comprising subjecting a material derived from nervous tissue a purification step selected from the group consisting of
A) extracting said material with an organic solvent,
B) loading a suspension of said material on a silica gel column and eluting said column with a mixture of organic and aqueous solvents,
and a combination of steps A) and B).

2. The process according to Claim 1 wherein said nervous tissue is derived from bovine brain.

3. The process according to Claim 1 wherein step A) comprises subjecting said nervous tissue material an extraction with a solvent mixture including a chlorinated hydrocarbon to obtain a first raw phospholipid-comprising mixture, subjecting said first raw phospholipid-comprising mixture a further purification step, and isolating the further purified product.

4. The process according to Claim 3 wherein the solvent mixture used for extracting nervous tissue comprises 1,1,1-trichloroethane and acetone.

5. The process according to Claim 1 wherein the mixture of organic and aqueous solvents used for elution in step B) comprises a chlorinated hydrocarbon and at least one alcohol.

6. The process according to Claim 5 wherein said chlorinated hydrocarbon is chloroform.

7. The process according to Claim 5 wherein said alcohol is selected from the group consisting of methanol, ethanol, propanols, butanols and pentanols.

8. The process according to Claim 1 wherein said suspension of a phospholipid mixture has been prepared by a method comprising the steps of

a) extracting bovine brain to obtain a crude extract comprising phospholipids,

b) filtering said crude extract to obtain a filtrate,

c) adding a precipitating agent to said filtrate to obtain a precipitate,

d) isolating said precipitate to obtain said first raw phospholipid-comprising mixture,

e) purifying said first raw mixture to obtain a second phospholipid-comprising mixture,

f) suspending said second mixture in a liquid.

9. The process according to Claim 1 further comprising a step for recovering said phospholipid mixture from relevant eluted fractions in step B.

10. The process according to Claim 9 wherein said recovery is performed by vacuum drying.

11. The process according to Claim 1 for the preparation of a phospholipid mixture completely free from unconventional viral agents while maintaining the pharmacological properties.

12. The process according to Claim 3 which is performed at room temperature for an extended period of time.

13. A phospholipid mixture preparable by the process according to Claim 1 and comprising

at least two components selected from the group consisting of phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, plasmalogen choline, plasmalogen ethanolamine, plasmalogen serine, phosphatidic acid, diphosphoinositide and sphingomyelin.

14. The mixture according to Claim 13 which comprises 30-50% of phosphatidylcholine, 24-44% of phosphatidylethanolamine, 7-13% of phosphatidylserine and 11-27% of phosphatidyl-inositol and minor phospholipids, all percentages being calculated on the basis of weight.

15. A phospholipid preparable by the process according to Claim 1 and comprising a single phospholipid fraction.

16. The phospholipid according to Claim 15 wherein the fraction is phosphatidylserine.

17. The process according to Claim 1 wherein the infective components are associated with unconventional agents.

18. The process according to Claim 1 wherein the infective components are "slow viruses".

19. The process according to Claim 1 wherein the infective components are associated with bovine spongiform encephalopathy.

20. A method for treatment of pathologies of the central nervous system and degenerative pathologies also associated with immune dysfunctions comprising the administration of an effective amount of a phospholipid mixture prepared according to Claim 1 to a patient in need thereof.

21. A pharmaceutical composition comprising, as active component, a phospholipid mixture preparable by the process as claimed in Claim 1, together with a pharmaceutically acceptable excipient.

22. The pharmaceutical composition according to Claim 21 being adapted for administration by parenteral route.

23. The pharmaceutical composition according to Claim 21 being adapted for administration by oral route.

24. A method for the removal of infective, viral components from bovine brain material which comprises

a) subjecting the brain material to extraction with a mixture of organic solvents,

b) subjecting the solvent extract so formed to a first purification,

c) precipitating said first purified material by the addition of a precipitating agent,

d) isolating the material precipitated,

e) purifying the isolated material, and optionally

f) suspending said purified material in a suspending liquid comprising a chlorinated hydrocarbon,

g) loading the suspension so formed on a silica gel column,

h) eluting said column with a mixture comprising organic and aqueous solvents, and

i) isolating the product after step e) or step h).

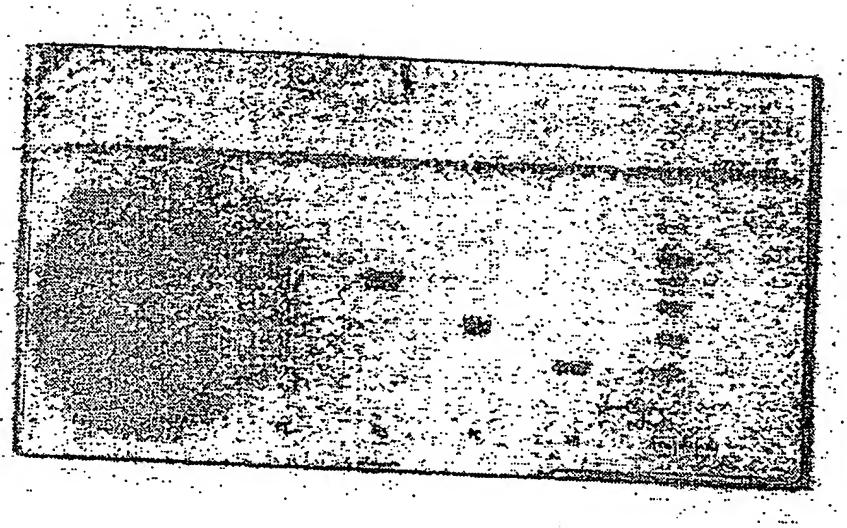

FIGURE 1

INTERNATIONAL SEARCH REPORT

International Application No PCT/EP 93/00915

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)[6]

According to International Patent Classification (IPC) or to both National Classification and IPC

Int.Cl. 5 C07F9/10; A61K31/685; A61K35/30

II. FIELDS SEARCHED

Minimum Documentation Searched[7]

Classification System	Classification Symbols
Int.Cl. 5	C07F ; A61K

Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched[8]

III. DOCUMENTS CONSIDERED TO BE RELEVANT[9]

Category °	Citation of Document,[11] with indication, where appropriate, of the relevant passages[12]	Relevant to Claim No.[13]
A	WO,A,9 107 417 (FIDIA S.P.A.) 30 May 1991 see page 15, line 30 - page 16, line 13	1-24
A	CHEMICAL ABSTRACTS, vol. 101, no. 22, 26 November 1984, Columbus, Ohio, US; abstract no. 198172c, page 377 ; see abstract & SU,A,1 102 603 (CENTRAL INSTITUTE FOR ADVANCED TRAINING OF PHYSICIANS) 15 July 1984	1-24
A	EP,A,0 150 712 (BIOIBERICA, S.A.) 7 August 1985 see claims	1-24

-/--

° Special categories of cited documents :[10]

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report
23 JULY 1993	12.08.93
International Searching Authority	**Signature of Authorized Officer**
EUROPEAN PATENT OFFICE	RYCKEBOSCH A.O.

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	EP,A,0 148 045 (FIDIA S.P.A.) 10 July 1985 see page 6, line 31 - page 7, line 4; claims	1-24

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 93/00915

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Arucle 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Remark: Although claims 20 is directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged effects of the compound/composition.

2. [ ] Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. [ ] As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.

EP 9300915
SA 73983

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report.
The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 23/07/93

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9107417	30-05-91	AU-A-	6727590	13-06-91
		CN-A-	1053067	17-07-91
		EP-A-	0454818	06-11-91
		JP-T-	4503076	04-06-92
EP-A-0150712	07-08-85	None		
EP-A-0148045	10-07-85	AU-A-	3563084	30-05-85
		BE-A-	901074	17-05-85
		CH-A-	661734	14-08-87
		DE-A-	3472406	04-08-88
		FR-A,B	2555050	24-05-85
		JP-A-	60132921	16-07-85
		LU-A-	85639	04-06-85
		US-A-	4595680	17-06-86